

NK CELLS ENGINEERING USING ADVANCED LENTIVIRAL VECTORS QUALITY AND DESIGN

THERAPEUTICS

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New challenge for Immunotherapy

The emergence of targeted immunotherapy, especially chimeric antigen receptor (CAR) T-cell therapy, has opened new possibilities, demonstrating tremendous success for patients with lymphoblastic leukemia. Natural Killer (NK) cells are other key immune effector cells which, contrary to T cells, do not require antigen priming and are at a low risk of Graft-Versus-Host Disease (GVHD), therefore offering the potential of an allogenic "offthe-shelf" therapeutic product. Nevertheless, primary NK cells exhibit a high resistance to lentiviral transduction, hampering transgene expression and consequently the generation of **CAR-NK** cells.

Pre-clinical and clinical CAR-NK cells manufacturing requires to be



performed using the same process. Here we show a manufacturing process of highly purified and concentrated third generation lentiviral vectors (LVs), available for a continuum going from Discovery to Clinic phases.

Integrating a control plan, allowing for the development and for the good manufacturing practices (GMP) production of custom lentiviral batches dedicated to clinical applications. Human primary NK cells, previously activated by artificial antigen-presenting cells (APC) are successfully transduced with such highly purified and concentrated Lentiviral vectors prior to be evaluated by in vitro cytotoxic assays and in vivo engraftments into mouse model.

C. Ex vivo Transduction of primary hNK cells

hNK CD56+ CD3- Cord blood cells were cultivated using the G-Rex® system (Wilsonwolf), in presence of IL-2, IL-15 and IL-21 and activated with artificial APCs. hNK cells were transduced twice at MOI 50 with a highly purified and concentrated Lentiviral vectors ILV-EF1-ZsGreen:



D. Engraftment of stably transduced hNK cells

hNK cells modified *ex vivo* are efficiently maintained *in vivo for* up to one month post engraftment : After activation with artificial APCs, NK CD56+ CD3- Cord blood cells were transduced twice at MOI 50 and then reactivated again prior to injection into NOG-hL15 mice (Taconic). PBMC are analysed:







Transduced human primary NK cells can efficiently kill HL60 and MOLM14 target cells in vitro (17h incubation at various Effector: Target cells ratio):





Ε. **CONCLUSIONS**

In this collaborative work, we show that the use of highly purified and concentrated self-inactivating lentiviral vectors in combination with an optimized transduction protocol, allows up to 65% of transduced human cord blood derived NK cells. Additionally, we show that transduction does not lead to viability nor phenotypic alterations of the transduced NK cells. Our approach not only achieves high transduction efficiency, leading to strong and stable transgene expression, but also **preserves** the **cytotoxic function** of the **NK cells**, *in vitro* and *in vivo*.

Many obstacles exist for clinical development of a CAR-based cellular therapy product, which requires efficient and safe delivery technologies, as well as gene expression level and duration tailoring. It's **possible to achieve** this, through the

use of delivery tools, which allow highly efficient gene transfer while maintaining transduced cell viability and phenotype. Here we propose a novel method allowing for the generation and production of **lentiviral vector engineered primary NK cells**, thus circumventing the problem of poor autologous CAR-T cell efficiency and gamma retrovirus associated risks.

This work lays the groundwork for novel cellular therapies based on lentivirally transduced primary NK cells. All these factors, as well as the ability to produce lentiviral vectors using Flash Therapeutics' GMP compliant production **platform**, offer additional safety considerations for clinical development and human use.



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